Analysis of Soil and eDNA from the Rhizosphere of a Quercus virginiana

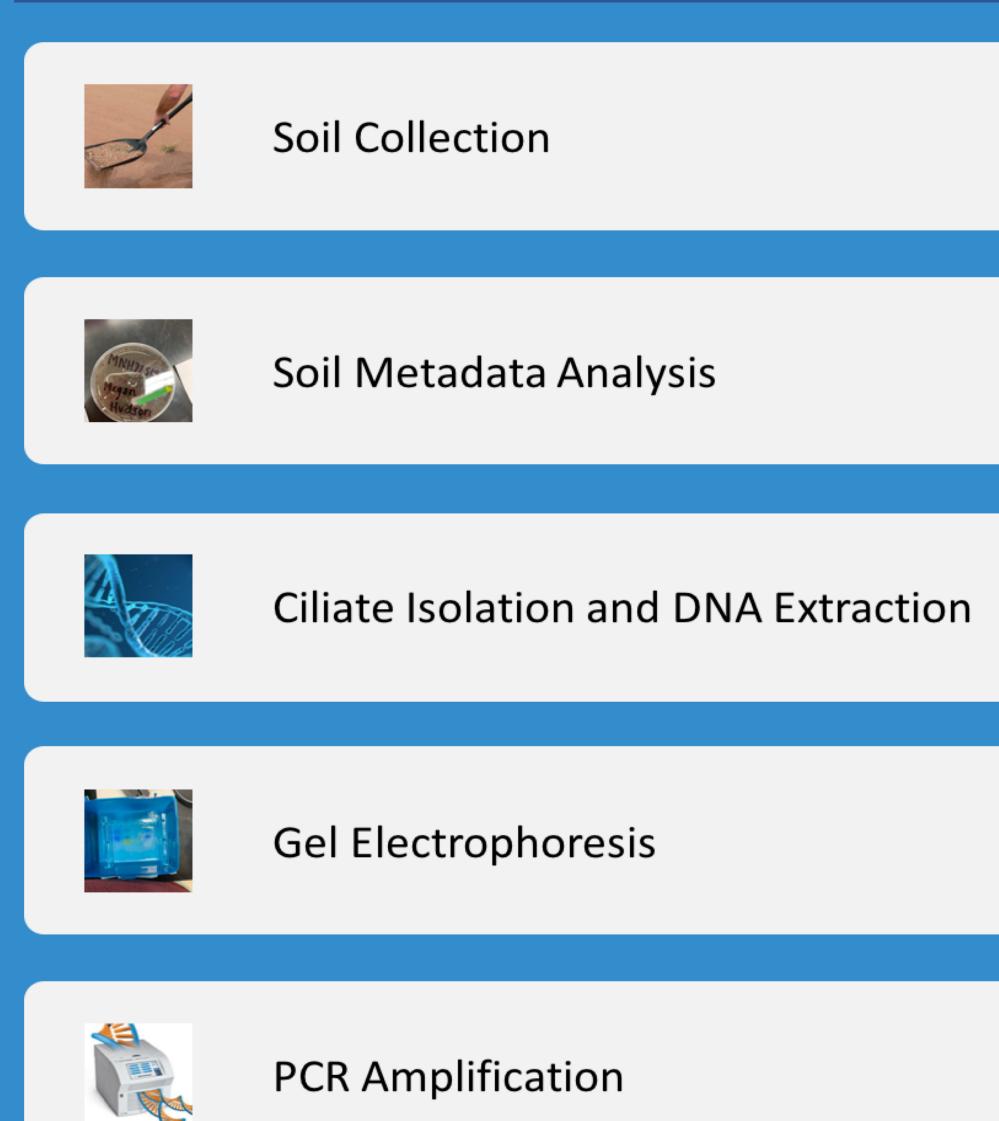
Introduction

- Ciliates are eukaryotic microorganisms that fall under one of the most controversial and diverse monophyletic supergroups, SAR [3].
- Soil microbiomes are largely unexplored and there is limited information regarding the connection between the biodiversity of ecosystems and the tree rhizosphere [2].
- The purpose of this experiment was to analyze the soil metadata collected from the Quercus virginiana, hypothesize how this connects to the overall community profile of the tree, and how organisms are impacted by the biodiversity.
- An eDNA sample from the rhizosphere of a *Quercus virginiana* was obtained due to the natural ability for ciliates to thrive in the rhizosphere [4].
- Methods were adapted through the use of silica beads in aims to isolate the eukaryotic 18SV4 region of terrestrial ciliates [2].
- The 18SV4 region was specifically chosen in attempts to analyze eukaryotic biodiversity and perform metabarcoding analysis.

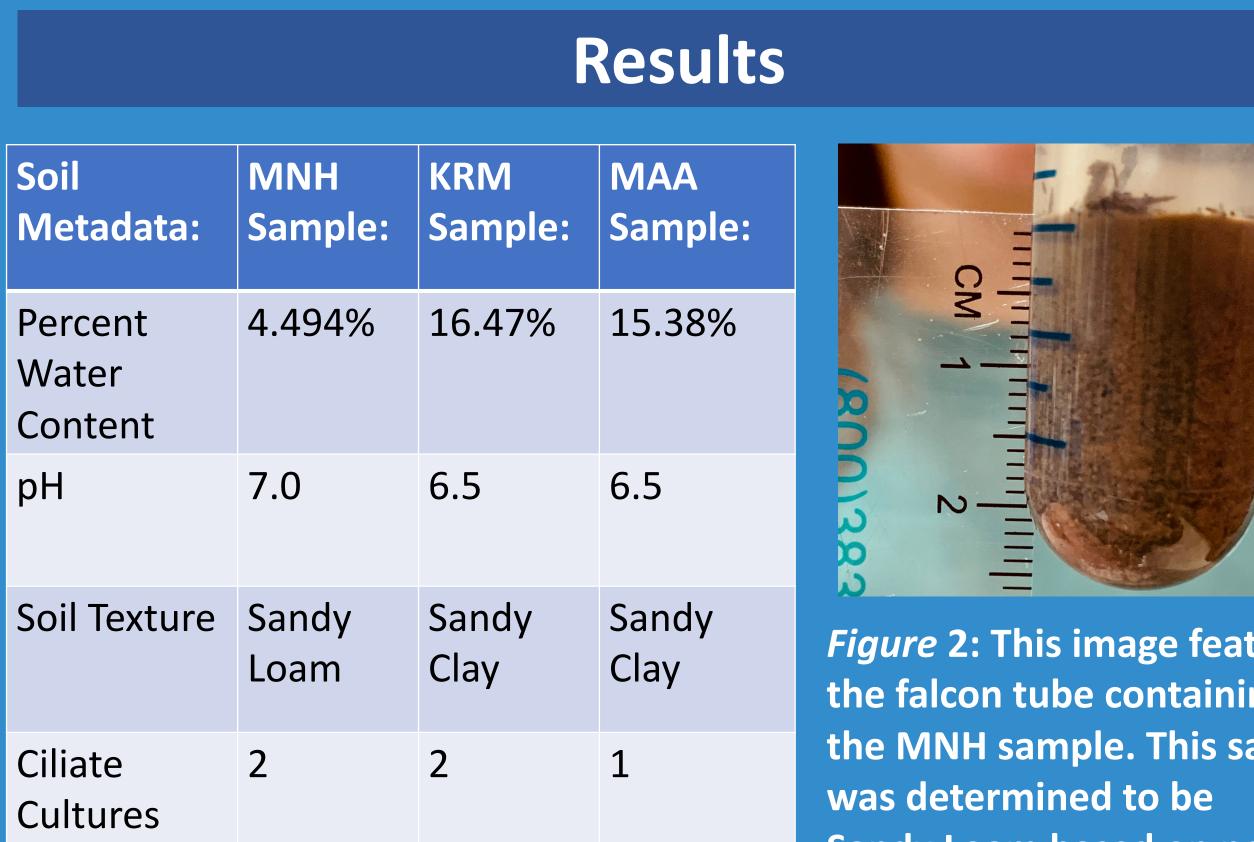


Figure 1: The soil samples were collected from this Quercus virginiana in the Vara Martin Daniel Plaza on **Baylor University's** campus.

Methods



Madison Ambrose, Megan Hudson, Kelsi Menzie



Tree and Leaf Metadata	
Tree Identification:	Que
Shape and Arrangement	Line
Margin	Enti
Venation	Pinn



Figure 4: Collected leaf under Figure 3: Leaves collected from a **Quercus virginiana** in the Vara Martin microscope to better distinguish Daniel Plaza. leaf metadata.

Sample	Concentration	A260/280:	A260/320:
MNH	33 ng/μL	1.3	0.05
KRM	326.3ng/µl	1.44	0.52

1000bp→

 $500bp \rightarrow$ $400bp \rightarrow$

200bp→

6

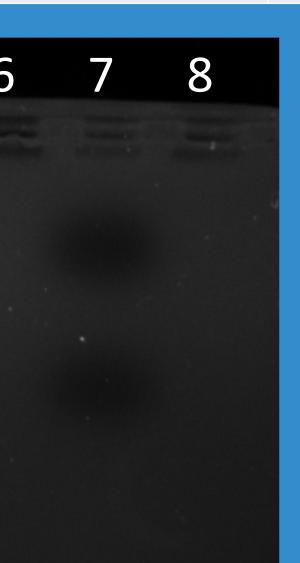
Figure 5: The soil sample was collected from the rhizosphere, near the shallow root of a tree. The DNA was extracted using a lysate method involving charcoal and silica beads. The DNA was then analyzed using a 1.5% agarose gel for electrophoresis; Lane 1: DNA Sample LL, Lane 2: blank, Lane 3: 5,000ng mass standard, Lane 4: blank, Lane 5: 15 ng mass standard, Lane 6: blank, Lane 7: DNA Sample MNH, Lane 8: blank. The image was taken by the MCB C305.

Figure 2: This image features the falcon tube containing the MNH sample. This sample Sandy Loam based on percent composition.

ercus virginiana

- nate







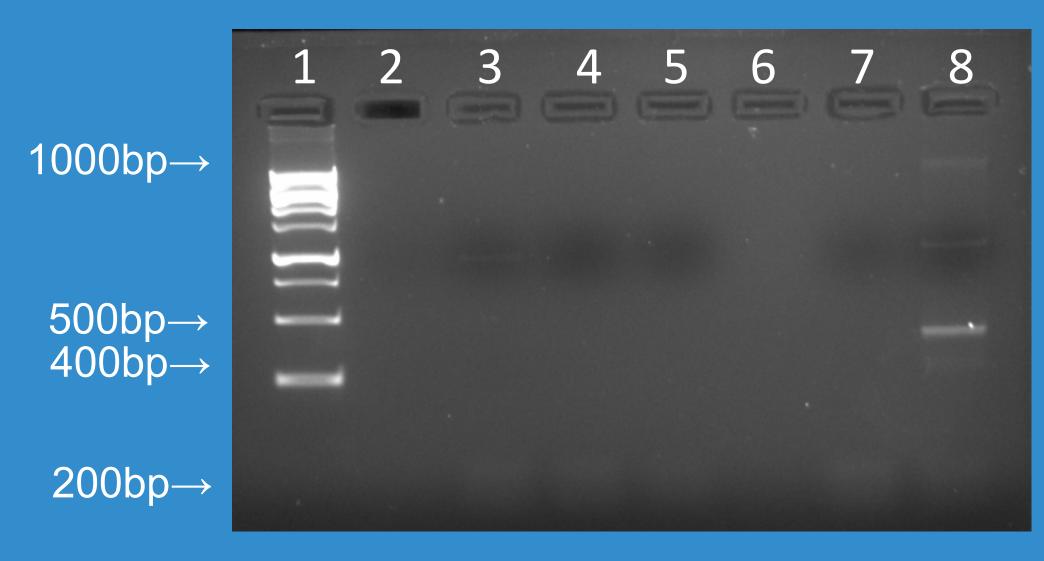


Figure 6: The DNA was prepared using the 2x Master Mix and the 18S V4 primer. The DNA was then analyzed using a 1.5% agarose gel for electrophoresis. Lanes from left to right, Lane 1: 5ul 1 kb ladder (company unknown), Lane 2: MM control, Lane 3: MM DNA, Lane 4: KRM control, Lane 5: KRM treatment, Lane 6: blank, Lane 7: LL control, Lane 8: LL DNA. The image was taken by the MCB C305.

Conclusion and Discussion

- eDNA.
- contain quantifiable eukaryotic eDNA.

- future studies.

Baylor University, CILI-CURE Dr. Tamarah Adair, Aadil Sheikh, Kaitlyn Armijo

- and Molecular Data. BioEssays, 40(4), 1700198.
- anoxic water. Molecular Ecology, 19, 21-31.
- 6. Tress of Texas ID by Leaf. (2019). Texas A&M Forest System.



Both MNH and KRM yielded an A260/280 concentration close to 1.8, indicating that there was a presence of

Results from PCR amplification of the KMR and MNH samples indicate that soil collected from the rhizosphere surrounding the *Quercus virginiana* did not

• Before eDNA was collected, ciliates were observed within the non-flooded plate sample. This could indicate that the V4 region of the ciliates could have been corrupted. This field experiment allowed research to be conducted analyzing soil metadata of the collected soil from the **Quercus Virginiana** rhizosphere and how this may affect the overall community profile of Live Oak trees. • Although the results from both samples were negative, adapted methods from A Rapid and Economical Method for Efficient DNA Extraction from Diverse Soils Suitable for Metagenomic Applications [1] produced positive DNA results for silica bead extraction and can be used in

Acknowledgments

References

1. Devi, S. G., Fathima, A. A., Radha, S., Arunraj, R., Curtis, W. R., & Ramya, M. (2015, July 13). A Rapid and Economical Method for Efficient DNA Extraction from Diverse Soils Suitable for Metagenomic Applications. 2. Foissner, W., & Stoeck, T. (2009). Morphological and Molecular Characterization of a New Protist Family, Sandmanniellidae n. fam. (Ciliophora, Colpodea), with Description of Sandmanniell terricolan. g., n. sp. from the Chobe Floodplain in Botswana. Journal of Eukaryotic Microbiology, 56(5), 472-483. 3. Grattepanche, J., Walker, L. M., Ott, B. M., Pinto, D. L., Delwiche, C. F., Lane, C. E., & Katz, L. A. (2018). Microbial Diversity in the Eukaryotic SAR Clade: Illuminating the Darkness Between Morphology 4. Ingham, E. R. (n.d.). Natural Resources Conservation Service. United States Department of Agriculture. 5. Stoeck, T., Bass, D., Nebel, M., Christen, R., Jones, M. D., Breiner, H., & Richards, T. A. (2010). Multiple marker parallel tag environmental DNA sequencing reveals a highly complex eukaryotic community in marine